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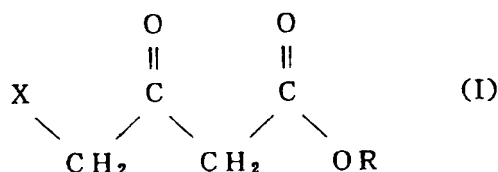
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D-81925 München (DE)(54) **Processes for production of optically active 4-halo-3-hydroxybutyric acid esters.**

(57) A microorganism that is capable of acting a 4-halo-acetoacetic acid ester shown by the general formula:



wherein X represents a halogen atom and R represents an optionally substituted alkyl group, alkenyl group, cycloalkyl group or aryl group, to produce an optically active 4-halo-3-hydroxybutyric acid ester or a preparation thereof is permitted to act on said 4-halo-acetoacetic acid ester and the product optically active 4-halo-3-hydroxybutyric acid ester is harvested. Thus, the desired optically active 4-halo-3-hydroxybutyric acid ester of high optical purity can be produced with commercial efficiency.

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FIELD OF THE INVENTION

The present invention relates to a process for producing an optically active 4-halo-3-hydroxybutyric acid ester characterized by permitting a microorganism or a preparation thereof to act on a 4-halo-3-acetoacetic acid ester and harvesting the product optically active 4-halo-3-hydroxybutyric acid ester.

BACKGROUND OF THE INVENTION

Optically active 4-halo-3-hydroxybutyric acid esters are important intermediates for the synthesis of various medicinal compounds, optically active biologically active substances and derivatives thereof.

For the production of an optically active 4-halo-3-hydroxybutyric acid ester, there is known an asymmetric enzymatic reduction (J. Am. Chem. Soc., 105, 5925 (1988); Ann. N. Y. Acad. Sci., 613, 628 (1990); Japanese Patent Application Laid-open No. 277494/1989; etc.), as well as an asymmetric reduction with the aid of a microorganism (Japanese Patent Application Laid-open No. 146191/1986, etc.) and an asymmetric reduction with the aid of *Lactobacillus fermentum* and *Lactobacillus kelfa* (Biocatalysis, Vol. 5, pp. 325 to 332 (1992)).

The enzymatic asymmetric reduction process, however, is disadvantageous in that the process is complicated and can not be carried out with simplicity. The asymmetric reduction processes with the aid of a microorganism are also disadvantageous in that the practically sufficient efficiency of the processes and the optical purity of the product optically active compound can not be obtained and that the microorganisms usable in the reactions are extremely restricted.

Under the circumstances, the establishment of an economical and expedient process for production of an optically active 4-halo-3-hydroxybutyric acid ester of high optical purity has been demanded.

SUMMARY OF THE INVENTION

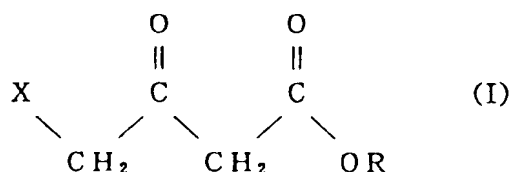
Accordingly, it is an object of the present invention to provide a process for producing an optically active 4-halo-3-hydroxybutyric acid ester of high optical purity expediently and efficiently with the aid of a microorganism.

It is another object of the invention to provide a commercially useful process for producing an optically active 4-halo-3-hydroxybutyric acid ester.

It is a further object of the invention to provide an efficient process for producing a (R)-4-halo-3-hydroxybutyric acid ester or a (S)-4-halo-3-hydroxybutyric acid ester with the aid of a microorganism.

The present inventors were interested in the utilization of a 4-halo-acetoacetic acid ester as a raw material with the aid of a microorganism for the economical and expedient production of an optically active 4-halo-3-hydroxybutyric acid ester of high optical purity. As a consequence, they discovered that certain strains selected from certain genera and species of microorganisms unexpectedly act on a 4-halo-acetoacetic acid ester to produce either a (R)-4-halo-3-hydroxybutyric acid ester or a (S)-4-halo-3-hydroxybutyric acid ester with a high optical purity and high reaction rates. The present invention has been accomplished on the basis of the above finding.

Thus, present invention provides a method of producing an optically active 4-halo-3-hydroxybutyric acid ester comprising permitting a microorganism or a preparation thereof to act on a 4-halo-acetoacetic acid ester shown by the general formula:



wherein X represents a halogen atom and R represents an optionally substituted alkyl group, alkenyl group, cycloalkyl group or aryl group, and harvesting the product optically active 4-halo-3-hydroxybutyric acid ester.

The microorganisms to be employed in accordance with the invention may be any strain of microorganism that is able to act on a 4-halo-acetoacetic acid ester to produce either a (S)-4-halo-3-hydroxybutyric acid ester or a (R)-4-halo-3-hydroxybutyric acid ester.

The microorganisms which is capable of producing a (S)-4-halo-3-hydroxybutyric acid ester include a strain of microorganism belonging to the genus *Brevibacterium*, the genus *Escherichia*, the genus *Kluyveromyces*, the genus *Leucosporidium*, the genus *Leuconostoc*, the genus *Lodderomyces*, the genus *Oosporidium*, the genus *Pediococcus*, the genus *Pityrosporum*, the genus *Rhodospiridium*, the genus *Sporidiobolus*, the genus *Stephanoascus*, the genus *Streptococcus*, the genus *Saccharomycopsis*, the genus *Wickerhamia*, the genus *Zygosaccharomyces*, the genus *Zygoascus*, *Lactobacillus buchneri*, *Lactobacillus bulgaricus*, *Lactobacillus casei*, *Lactobacillus delbrueckii*, *Lactobacillus frigidus*, *Lactobacillus hilgardii*, *Lactobacillus lactis*, *Lactobacillus malefermentans*, *Lactobacillus plantarum* and *Lactobacillus xylosus*.

The microorganisms which produce (R)-4-halo-3-hydroxybutyric acid ester include a strain of microorganism belonging to the genus *Arthrobacter*, the genus *Leuconostoc*, the genus *Streptococcus*, the genus *Sporolactobacillus*, *Lactobacillus brevis*, *Lactobacillus collinoides*, *Lactobacillus leichmannii* and *Lactobacillus viridescens*.

Such a microorganism is generally grown in a culture medium and, then, submitted to the reaction with a 4-halo-acetoacetic acid ester. A preparation of such microorganism may instead be used in the reaction with a 4-halo-acetoacetic acid ester.

DETAILED DESCRIPTION OF THE INVENTION

As the halogen atom represented by X in the 4-halo-acetoacetic acid ester shown by the formula (I) as used in the present invention, there may be mentioned chlorine, bromine, iodine and so on.

Examples of the alkyl group represented by R include a straight chain or branched alkyl group having 1 to 8 carbon atoms such as methyl group, ethyl group, n-propyl group, isopropyl group, n-butyl group, i-butyl group, tert-butyl group, n-pentyl group, tert-pentyl group, hexyl group, heptyl group and octyl group.

As the alkenyl group represented by R, there may be mentioned a straight chain or branched alkenyl group having 2 to 6 carbon atoms such as vinyl group, propenyl group and 2-butenyl group.

The cycloalkyl group is exemplified as a monocycloalkyl group having 3 to 8 carbon atoms such as cyclopropyl group, cyclobutyl group, cyclopentyl group, cyclohexyl group and cycloheptyl group.

Examples of the aryl group include an aryl group having 6 to 14 carbon atoms such as phenyl group and naphthyl group.

The alkyl group, alkenyl group, cycloalkyl group or aryl group represented by R may optionally be substituted with a substituent.

As such a substituent, any one can be employed as far as the reaction is not adversely affected, thus including substituents generally employed for those groups.

Typical examples of these substituent include a halogen atom such as iodine, chlorine, fluorine and bromine, nitro group, an alkoxy group having 1 to 4 carbon atoms (for example methoxy group, ethoxy group, propoxy group, butoxy group, etc.), an aryl group having 6 to 14 carbon atoms (e.g. phenyl group, naphthyl group and the like), an alkyl group having 1 to 8 carbon atoms (for instance, methyl group, ethyl group, n-propyl group, i-propyl group, n-butyl group, i-butyl group, tert-butyl group, octyl group, etc.), a cycloalkyl group having 3 to 8 carbon atoms (e.g. cyclopentyl group, cyclohexyl group, etc.) and so on. The number of the substituents may preferably be 1 to 4.

Practically preferred examples of the group represented by R may include a straight chain or branched alkyl group having 1 to 4 carbon atoms such as methyl group, ethyl group, n-propyl group and i-propyl group, and an optionally substituted phenyl group (e.g. phenyl group, tolyl group, and the like).

The microorganisms to be employed in accordance with the invention may be any strain of microorganism that is able to act on a 4-halo-acetoacetic acid ester to produce a (S)-4-halo-3-hydroxybutyric acid ester or a (R)-4-halo-3-hydroxybutyric acid ester.

The examples of those microorganisms which are able to act on a 4-halo-acetoacetic acid ester to produce a (S)-4-halo-3-hydroxybutyric acid ester include, among others, the genus *Brevibacterium*, the genus *Escherichia*, the genus *Kluyveromyces*, the genus *Leucosporidium*, the genus *Leuconostoc*, the genus *Lodderomyces*, the genus *Oosporidium*, the genus *Pediococcus*, the genus *Pityrosporum*, the genus *Rhodospiridium*, the genus *Sporidiobolus*, the genus *Stephanoascus*, the genus *Streptococcus*, the genus *Saccharomycopsis*, the genus *Wickerhamia*, the genus *Zygosaccharomyces*, the genus *Zygoascus*, *Lactobacillus buchneri*, *Lactobacillus bulgaricus*, *Lactobacillus casei*, *Lactobacillus delbrueckii*, *Lactobacillus frigidus*, *Lactobacillus hilgardii*, *Lactobacillus lactis*, *Lactobacillus malefermentans*, *Lactobacillus plantarum* and *Lactobacillus xylosus*.

As typical examples of the strain of microorganism that is able to act on a 4-halo-3-acetoacetic acid ester to produce a (S)-4-halo-3-hydroxybutyric acid ester, there may be mentioned

the genus *Escherichia*: *Escherichia coli* IFO 3302, etc.,
 the genus *Brevibacterium*: *Brevibacterium ammoniagenes* IFO 12072, etc.,
 the genus *Kluyveromyces*: *Kluyveromyces lactis* IFO 1267, etc.,
 the genus *Leucosporidium*: *Leucosporidium scottii* IFO 1924, etc.,
 5 the genus *Leuconostoc*: *Leuconostoc dextranicum* IFO 3347, etc.,
 the genus *Lodderomyces*: *Lodderomyces elongisporus* IFO 1676, etc.,
 the genus *Oosporidium*: *Oosporidium margartieerum* IFO 1208, etc.,
 the genus *Pediococcus*: *Pediococcus parvulus* IFO 12233, etc.,
 the genus *Pityrosporum*: *Pityrosporum ovale* IFO 0656, etc.,
 10 the genus *Rhodospiridium*: *Rhodospiridium diobovatum* IFO 1830, etc.,
 the genus *Sporidiobolus*: *Sporidiobolus pararoseus* IFO 0376, etc.,
 the genus *Stephanoascus*: *Stephanoascus ciferrii* IFO 1854, etc.,
 the genus *Streptococcus*: *Streptococcus equi* NRIC 1138, etc.,
 the genus *Saccharomycopsis*: *Saccharomycopsis capsularis* DSM 70560, etc.,
 15 the genus *Wickerhamia*: *Wickerhamia fivorescens* DSM 70715, etc.,
 the genus *Zygosaccharomyces*: *Zygosaccharomyces bailii* DSM 70492, etc.,
 the genus *Zygoascus*: *Zygoascus hellenicus* IFO 1575, etc.,
Lactobacillus buchneri: *Lactobacillus buchneri* NRIC 1040, etc.,
Lactobacillus bulgaricus: *Lactobacillus bulgaricus* NRIC 1041, etc.,
 20 *Lactobacillus casei*: *Lactobacillus casei* NRIC 1044, etc.,
Lactobacillus delbrueckii: *Lactobacillus delbrueckii* IAM 1085, etc.,
Lactobacillus frigidus: *Lactobacillus frigidus* NRIC 1079, etc.,
Lactobacillus hilgardii: *Lactobacillus hilgardii* NRIC 1060, etc.,
Lactobacillus lactis: *Lactobacillus lactis* DSM 20073, etc.,
 25 *Lactobacillus malefermentans*: *Lactobacillus malefermentans* NRIC 1081, etc.,
Lactobacillus plantarum: *Lactobacillus plantarum* IFO 3070, etc.,
Lactobacillus xylosus: *Lactobacillus xylosus* NRIC 1074, etc. and the like.

The examples of those microorganisms which has capacity of acting on a 4-halo-acetoacetic acid ester to produce a (R)-4-halo-3-hydroxybutyric acid ester include those belong to the genus *Arthrobacter*, the
 30 genus *Leuconostoc*, the genus *Streptococcus*, the genus *Sporolactobacillus*, *Lactobacillus brevis*, *Lactobacillus collinoides*, *Lactobacillus leichmannii* and *Lactobacillus viridescens*.

Practical examples of the strain of microorganism that is able to act on a 4-halo-acetoacetic acid ester to produce a (R)-4-halo-3-hydroxybutyric acid ester include

the genus *Arthrobacter*: *Arthrobacter giobiformis* IFO 12140, etc.,
 35 the genus *Leuconostoc*: *Leuconostoc dextranicum* ATCC 17072, etc.,
 the genus *Streptococcus*: *Streptococcus faecalis* IFO 12964, *Streptococcus faecium* NRIC 1145, *Streptococcus* sp. IFO 3535, etc.,
 the genus *Sporolactobacillus*: *Sporolactobacillus inulinus* TUA 343, etc.,
Lactobacillus brevis: *Lactobacillus brevis* NRIC 1037, etc.,
 40 *Lactobacillus collinoides*: *Lactobacillus collinoides* NRIC 1049, etc.,
Lactobacillus leichmannii: *Lactobacillus leichmannii* JCM 1557, etc.,
Lactobacillus viridescens: *Lactobacillus viridescens* NRIC 1073, etc. and the like.
 At least one strain of microorganism among them can be employed.

For the purposes of the invention, any of wild strains, mutants and recombinant strains which can be
 45 obtained by a genetic engineering technique such as cell fusion or gene manipulation, that is able to act on a 4-halo-acetoacetic acid ester to produce an optically active 4-halo-3-hydroxybutyric acid ester can be advantageously employed.

The microorganisms identified hereinabove by IFO numbers are described in the "List of Cultures Ed. 8, Vol. 1 (1988)" published by Institute for Fermentation, Osaka (IFO), Japan and are available from the
 50 same Institute. The microorganisms designated by JCM numbers are listed in "Catalogs of Microbial Strains Ed. 4 (1989)" published by the Culture Collection of The Institute of Physical and Chemical Research, Japan and available from the same Culture Collection. The microorganisms designated by ATCC numbers are listed in "Catalogue of Bacteria Phages rDNA Vectors, Ed. 16 (1985)" and "Catalogue of Fungi/Yeast, Ed. 17 (1987)" each published by the American Type Culture Collection (ATCC) and are
 55 available from the same organization. The microorganisms designated by DSM numbers are listed in "Catalogue of Strains (1983)" of Deutsch Sammlung von Mikroorganismen (DSM) and are available from the same organization. The microorganisms designated by IAM numbers are available from Institute for Applied Microbiology of Tokyo University and the microorganisms designated by NRIC numbers and TUA

numbers are available from Tokyo Agricultural University.

A microorganism, such as the above, is usually grown in a culture medium and, then, submitted to the reaction with a 4-halo-acetoacetic acid ester.

The medium which is used for growing the strain for use in the invention is not critical in composition only if the selected strain may grow and multiply therein. The medium is generally a fluid medium containing sources of carbon and nitrogen and other nutrients. Any carbon source which the strain can utilize may be employed. As the sources of carbon, there may be employed various carbohydrates such as glucose, fructose, sucrose, dextrin, starch, etc., alcohols such as sorbitol, methanol, ethanol, glycerol, etc., organic acids such as fumaric acid, citric acid, acetic acid, propionic acid, etc. and the corresponding salts, hydrocarbons such as paraffin, and various mixtures thereof. The sources of nitrogen include, among others, inorganic acid ammonium salts such as ammonium chloride, ammonium sulfate, ammonium phosphate, etc., organic acid ammonium salts such as ammonium fumarate, ammonium citrate, etc., inorganic or organic nitrogenous materials such as meat extract, yeast extract, malt extract, peptone (polypeptone), corn steep liquor, casein hydrolysate, urea, etc., and various mixtures thereof.

In the medium, there may be incorporated appropriate amounts of those nutrients which are commonly employed in the cultivation of microorganisms, such as inorganic salts, trace metal salts and vitamins. Where necessary, there may also be incorporated factors which may promote growth of the strain used and/or factors which may augment its ability to produce the object compound of the invention, such as a 4-halo-acetoacetic acid ester, as well as a buffer substance which may assist in the maintenance of the medium at a given pH.

The cultivation of the microorganism is carried out under conditions optimal for the growth of the particular strain, for example at a medium pH in the range of about 3.0 to 9.5, preferably about 4 to 8, and an incubation temperature in the range of about 20 to 45 °C, preferably about 25 to 37 °C. The cultivation may be aerobic or anaerobic. The cultivation time may, for example, be 5 to 120 hours, preferably about 12 to 72 hours.

The desired optically active 4-halo-3-hydroxybutyric acid ester can be produced as a 4-halo-acetoacetic acid ester is added to a dispersion of cells of the microorganism or a preparation thereof for asymmetric reduction.

The method of production of an optically active 4-halo-3-hydroxybutyric acid ester from the corresponding 4-halo-acetoacetic acid ester may, for example, be whichever of the following alternatives: (1) the method which comprises adding a 4-halo-acetoacetic acid ester to a culture broth as such, (2) the method which comprises separating the microbial cells from the culture broth, e.g. by centrifugation, resuspending the cells, either as they are or after washing with water, in a buffer solution, water or the like, and adding a 4-halo-acetoacetic acid ester to the resulting cell suspension, (3) the method which comprises using treated preparation of cells such as disrupted cells, acetone-treated cells, lyophilized cells and so on and adding the material to the resulting cell preparation, and (4) the method which comprises immobilizing these cells or preparations thereof and adding the material thereto. There are cases in which this reaction proceeds with advantage in the presence of a carbon source, such as glucose, sucrose, methanol or ethanol, which serves as an energy source.

The optimal cell concentration of the reaction system cannot be stated in general terms, for it is significantly dependent on the species or strain of microorganism employed. However, the concentration should be in the range where the efficiency of leaving the desired optically active compound intact will not be adversely affected. A typical cell concentration may for example be, on a dry cell basis, about 0.1 to 100 g/liter and preferably about 1 to 50 g/liter.

The cells may be wet viable cells or any preparation thereof, such as disrupted cells, acetone-treated cells, lyophilized cells and so on. These cells or cell preparations may be immobilized by known techniques such as the polyacrylamide gel method, sulfur-containing polysaccharide gel method (e.g. carrageenin gel method), alginic acid gel method, agar gel method and so on. The enzyme purified from such a cell preparation can also be employed. The enzyme can be obtained by using known purification processes in a suitable combination.

The corresponding 4-halo-acetoacetic acid ester can be used as it is or in the form of a solution in water or an organic solvent which will not interfere with the reaction or a dispersion prepared with a surfactant. The 4-halo-acetoacetic acid ester may be added in bolus at the beginning of the reaction or in several installments.

The reaction conditions can be selected from the ranges that will not detract from the yield of the object compound. For example, the pH of the reaction system can be selected from the range of pH about 3 to 10 and preferably pH about 5 to 9. The reaction temperature can be selected from the range of, for example, 10 to 60 °C and preferably from 20 to 40 °C. The reaction can be conducted with stirring or under stationary

conditions for about 1 to 120 hours. The concentration of a 4-halo-acetoacetic acid ester as the substrate is not particularly critical and is preferably about 0.1 to 20 weight % and more preferably about 0.2 to 10 weight %.

The optically active 4-halo-3-hydroxybutyric acid ester produced by the reaction can be harvested by the separation and purification procedures generally known. For example, the optically active 4-halo-3-hydroxybutyric acid ester can be easily obtained by subjecting the reaction mixture, directly or after separation of the cells, to the conventional purification procedure such as extraction with an organic solvent, distillation and column chromatography. The optical purity of optically active 4-halo-3-hydroxybutyric acid ester can be measured by high performance liquid chromatography (HPLC) using an optical resolution column.

Thus, according to the method of the present invention using a microorganism or preparation thereof, an optically active 4-halo-3-hydroxybutyric acid ester of high optical purity can be produced expediently and efficiently, therefore the method is commercially useful.

The following examples are intended to illustrate the invention in further detail and should by no means be construed as delimiting the scope of the invention.

EXAMPLES

In the examples, the quantitative determination of ethyl 4-chloro-3-hydroxybutyrate in reaction mixture was carried out by subjecting ethyl 4-chloro-3-hydroxybutyrate obtained by the reaction to gas chromatography using a column (column: Thermon 3000, Chromosorb W; length: 2 m; the column temperature: 140 °C). The optical purity determination thereof was carried out by subjecting the optically active 4-halo-3-hydroxybutyric acid ester to removing the solvent off, then directly to high performance liquid chromatography using an optical resolution column (column: Chiralpack AS, Daicel Chemical Industries, Ltd.; moving phase: n-hexane/isopropyl alcohol/ethanol/cyclohexanol = 92/2.5/1.25/0.25; wavelength: 220 nm; flow rate: 1 ml/min.). Under the above operating conditions, the retention time of ethyl 4-chloro-3-hydroxybutyrate was 13.8 minutes for (S) and 15.1 minutes for (R).

Example 1

A test tube of 21 mm in diameter was charged with 5 ml of the following growth medium and, after sterilization, was inoculated with one of the microbial stains shown in Tables 1 to 4. The inoculated tube was incubated under shaking at 30 °C for 48 hours. Subsequently cells were isolated by centrifuging to obtain viable cells.

(A) Growth medium for a yeast

Glucose	2.0 weight %
Yeast extract	0.3 weight %
Malt extract	0.3 weight %
Polypeptone	0.5 weight %
pH	6.0

(B) Growth medium for a bacterium

Glucose	2.0 weight %
Yeast extract	0.3 weight %
Meat extract	0.3 weight %
Polypeptone	0.5 weight %
Ammonium sulfate	0.2 weight %
Potassium primary phosphate	0.1 weight %
Magnesium sulfate	0.05 weight %
pH	7.0

(C) Growth medium for a lactic acid bacterium

Glucose	2.0 weight %
Yeast extract	1.0 weight %
Polypeptone	1.0 weight %
Sodium acetate	1.0 weight %
Magnesium sulfate	0.02 weight %
Manganese sulfate	1 ppm
Ferrous sulfate	1 ppm
Sodium chloride	1 ppm
Calcium carbonate	1 weight %
pH	6.8

Then, a test tube of 21 mm in diameter was charged with 2.5 ml of 0.1 M potassium phosphate buffer (pH 6.5) containing 125 mg of glucose and said viable cells were suspended therein. Afterwards, 25 μ l of ethyl 4-chloro-acetoacetate was added to the suspension and the reaction was conducted on a reciprocating shaker at 30 °C for 48 hours.

After completion of the reaction, 1 ml of the reaction suspension was extracted with 2 ml of ethyl acetate. The ethyl acetate extract was subjected to gas chromatography to determine the amount of the product ethyl 4-chloro-3-hydroxybutyrate.

Then, after the solvent was removed with use of a rotary evaporator to give a syrup. The syrup was dissolved in n-hexane and the absolute configuration and optical purity of the optically active ethyl 4-chloro-3-hydroxybutyrate were determined using high performance liquid chromatography. The results are set forth in Tables 1 to 4.

Table 1

Name of Microorganism	Product amount of ethyl 4-chloro-3-hydroxybutyrate (mg/ml)	Absolute configuration	Optical purity (% e.e.)
Brevibacterium ammoniagenes IFO 12072	3.0	S	96
Escherichia coli IFO 3302	0.9	S	82
Kluyveromyces lactis IFO 1267	7.7	S	99
Lactobacillus buchneri NRIC 1040	10.0	S	46
Lactobacillus bulgaricus NRIC 1041	2.7	S	90
Lactobacillus casei NRIC 1044	3.5	S	99
Lactobacillus delbrueckii IAM 1085	4.0	S	67
Lactobacillus frigidus NRIC 1079	5.1	S	72
Lactobacillus hilgardii NRIC 1060	7.8	S	92

Table 2

Name of Microorganism	Product amount of ethyl 4-chloro-3-hydroxybutyrate (mg/ml)	Absolute configuration	Optical purity (% e.e.)
Lactobacillus lactis DSM 20073	4.4	S	63
Lactobacillus malefermentans NRIC 1081	6.9	S	99
Lactobacillus plantarum IFO 3070	10.0	S	99
Lactobacillus xylosus NRIC 1074	7.3	S	95
Leuconostoc dextranicum IFO 3347	8.9	S	57
Leucosporidium scottii IFO 1924	7.1	S	37
Lodderomyces elongisporus IFO 1676	4.6	S	70
Oosporidium margartieerum IFO 1208	3.8	S	58
Pediococcus parvurus IFO 12233	6.2	S	83

Table 3

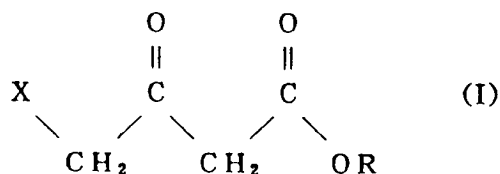
Name of Microorganism	Product amount of ethyl 4-chloro-3-hydroxybutyrate (mg/ml)	Absolute configuration	Optical purity (% e.e.)
Pityrosporum ovale IFO 0656	2.7	S	72
Rhodospiridium diobovatum IFO 1830	7.1	S	39
Saccharomycopsis capsularis DSM 70560	3.5	S	99
Sporidiobolus paraseus IFO 0376	3.4	S	48
Stephanoascus ciferrii IFO 1854	2.7	S	99
Streptococcus equi NRIC 1138	3.6	S	42
Wickerhamia fivorescens DSM 70715	4.5	S	40
Zygosaccharomyces bailii DSM 70492	3.5	S	66
Zygoascus hellenicus IFO 1575	3.5	S	67

Table 4

Name of Microorganism	Product amount of ethyl 4-chloro-3-hydroxybutyrate (mg/ml)	Absolute configuration	Optical purity (% e.e.)
Arthrobacter globiformis IFO 12140	2.1	R	70
Lactobacillus brevis NRIC 1037	2.1	R	86
Lactobacillus collinoides NRIC 1049	4.0	R	73
Lactobacillus leichmannii JCM 1557	3.2	R	89
Leuconostoc dextranicum ATCC 17072	2.8	R	94
Lactobacillus viridescens NRIC 1073	7.3	R	73
Sporolactobacillus inulinus TUA 343	1.5	R	49
Streptococcus faecalis IFO 12964	1.3	R	86
Streptococcus faecium NRIC 1145	1.6	R	82
Streptococcus sp. IFO 3535	2.1	R	92

Claims

1. A process for producing an optically active 4-halo-3-hydroxybutyric acid ester which comprises permitting a microorganism capable of acting on a 4-halo-acetoacetic acid ester shown by the general formula:

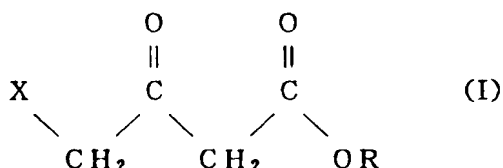


wherein X represents a halogen atom and R represents an optionally substituted alkyl group, alkenyl group, cycloalkyl group or aryl group, to produce an optically active 4-halo-3-hydroxybutyric acid ester or a preparation thereof to act on said 4-halo-acetoacetic acid ester and harvesting the product optically active 4-halo-3-hydroxybutyric acid ester.

2. The process for producing an optically active 4-halo-3-hydroxybutyric acid ester according to claim 1, which comprises permitting a microorganism that is capable of acting on a 4-halo-acetoacetic acid ester to selectively produce a (S)-4-halo-3-hydroxybutyric acid ester or a preparation thereof to act on the 4-halo-acetoacetic acid shown by the general formula (I) and harvesting the product (S)-4-halo-3-hydroxybutyric acid ester.
3. The process for producing an optically active 4-halo-3-hydroxybutyric acid ester according to claim 2, wherein said microorganism capable of producing a (S)-4-halo-3-hydroxybutyric acid ester is a strain of microorganism being selected from the group of microorganisms belonging to the genus *Brevibacterium*, the genus *Escherichia*, the genus *Kluyveromyces*, the genus *Leucosporidium*, the genus *Leuconostoc*, the genus *Lodderomyces*, the genus *Oosporidium*, the genus *Pediococcus*, the genus *Pityrosporum*, the genus *Rhodospiridium*, the genus *Sporidiobolus*, the genus *Stephanoascus*, the genus *Streptococcus*, the genus *Saccharomycopsis*, the genus *Wickerhamia*, the genus *Zygosaccharomyces* and the genus *Zygoascus*.
4. The process for producing an optically active 4-halo-3-hydroxybutyric acid ester according to claim 3, wherein said strain of microorganism is a strain of microorganism being selected from the group of microorganisms belonging to *Brevibacterium ammoniagenes*, *Escherichia coli*, *Kluyveromyces lactis*, *Leucosporidium scottii*, *Leuconostoc dextranicum*, *Lodderomyces elongisporus*, *Oosporidium margaritaeum*, *Pediococcus parvurus*, *Pityrosporum ovale*, *Rhodospiridium diobovatum*, *Sporidiobolus parroseus*, *Stephanoascus ciferrii*, *Streptococcus equi*, *Saccharomycopsis capsularis*, *Wickerhamia fivorescens*, *Zygosaccharomyces bailii* and *Zygoascus hellenicus*.
5. The process for producing an optically active 4-halo-3-hydroxybutyric acid ester according to claim 2, wherein said microorganism capable of producing a (S)-4-halo-3-hydroxybutyric acid ester is a strain of microorganism being selected from the group of microorganisms belonging to *Lactobacillus buchneri*, *Lactobacillus bulgaricus*, *Lactobacillus casei*, *Lactobacillus delbrueckii*, *Lactobacillus frigidus*, *Lactobacillus hilgardii*, *Lactobacillus lactis*, *Lactobacillus malefermentans*, *Lactobacillus plantarum* and *Lactobacillus xylosus*.
6. The process for producing an optically active 4-halo-3-hydroxybutyric acid ester according to claim 1, which comprises permitting a microorganism that is capable of acting on a 4-halo-acetoacetic acid ester to selectively produce a (R)-4-halo-3-hydroxybutyric acid ester or a preparation thereof to act on the 4-halo-acetoacetic acid shown by the general formula (I) and harvesting the product (R)-4-halo-3-hydroxybutyric acid ester.
7. The process for producing optically active 4-halo-3-hydroxybutyric acid ester according to claim 6, wherein said microorganism capable of producing a (R)-4-halo-3-hydroxybutyric acid ester is a strain of microorganism being selected from the group of microorganisms belonging to the genus *Arthrobacter*, the genus *Leuconostoc*, the genus *Streptococcus*, the genus *Sporolactobacillus*.
8. The process for producing an optically active 4-halo-3-hydroxybutyric acid ester according to claim 7, wherein said strain of microorganism is a strain of microorganism being selected from the group of

microorganisms belonging to *Arthrobacter giobiformis*, *Leuconostoc dextranicum*, *Streptococcus faecalis*, *Streptococcus faecium*, *Streptococcus* sp. IFO 3535 and *Sporolactobacillus inulinus*.

9. The process for producing optically active 4-halo-3-hydroxybutyric acid ester according to claim 6, wherein said microorganism capable of producing a (R)-4-halo-3-hydroxybutyric acid ester is a strain of microorganism being selected from the group of microorganisms belonging to *Lactobacillus brevis*, *Lactobacillus collinoides*, *Lactobacillus leichmannii* and *Lactobacillus viridescens*.
10. The process for producing an optically active 4-halo-3-hydroxybutyric acid ester according to claim 1, which comprises growing a microorganism in a fluid medium and adding said 4-halo-acetoacetic acid ester shown by the general formula (I) to a dispersion of said microorganism or a preparation thereof.
11. The process for producing an optically active 4-halo-3-hydroxybutyric acid ester according to claim 1, which comprises permitting said microorganism or preparation thereof to act on said 4-halo-acetoacetic acid ester shown by the general formula (I) at pH 3 to 10 and a temperature of 10 to 60 °C.
12. The process for producing an optically active 4-halo-3-hydroxybutyric acid ester according to claim 1, wherein the concentration of said 4-halo-acetoacetic acid ester shown by the general formula (I) is 0.1 to 20 weight %.
13. The process for producing an optically active 4-halo-3-hydroxybutyric acid ester according to claim 1, which comprises permitting said microorganism or preparation thereof to act on said 4-halo-acetoacetic acid ester shown by the general formula (I) in the presence of a carbon source.
14. A process for asymmetrically reducing a 4-halo-acetoacetic acid ester shown by the general formula:



wherein X represents a halogen atom and R represents an optionally substituted alkyl group, alkenyl group, cycloalkyl group or aryl group, which comprises:

subjecting said 4-halo-acetoacetic acid ester to a microorganism capable of acting on said 4-halo-acetoacetic acid ester to asymmetrically reduce into an optically active 4-halo-3-hydroxybutyric acid ester or a preparation derived therefrom, and

recovering the product which is an optically active 4-halo-3-hydroxybutyric acid ester.

15. The use of a microorganism strain being selected from the group of microorganisms belonging to the genus *Brevibacterium*, the genus *Escherichia*, the genus *Kluyveromyces*, the genus *Leucosporidium*, the genus *Leuconostoc*, the genus *Lodderomyces*, the genus *Oosporidium*, the genus *Pediococcus*, the genus *Pityrosporum*, the genus *Rhodospiridium*, the genus *Sporidiobolus*, the genus *Stephanoascus*, the genus *Streptococcus*, the genus *Saccharomycopsis*, the genus *Wickerhamia*, the genus *Zygosaccharomyces* and the genus *Zygoascus*, or from *Lactobacillus buchneri*, *Lactobacillus bulgaricus*, *Lactobacillus casei*, *Lactobacillus delbrueckii*, *Lactobacillus frigidus*, *Lactobacillus hilgardii*, *Lactobacillus lactis*, *Lactobacillus malefermentans*, *Lactobacillus plantarum* and *Lactobacillus xylosus* for the production of (S)-4-halo-3-hydroxybutyric acid ester.
16. The use of a microorganism strain being selected from the group of microorganisms belonging to the genus *Arthrobacter*, the genus *Leuconostoc*, the genus *Streptococcus*, the genus *Sporolactobacillus*, or from *Lactobacillus brevis*, *Lactobacillus collinoides*, *Lactobacillus leichmannii* and *Lactobacillus viridescens* for the production of (R)-4-halo-3-hydroxybutyric acid ester.

(19)



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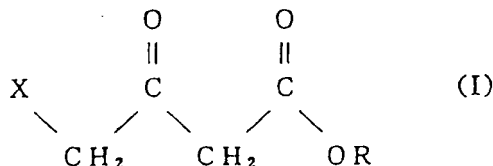
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(54) **Processes for production of optically active 4-halo-3-hydroxybutyric acid esters.**

(57) A microorganism that is capable of acting a 4-halo-acetoacetic acid ester shown by the general formula:

be produced with commercial efficiency.



wherein X represents a halogen atom and R represents an optionally substituted alkyl group, alkenyl group, cycloalkyl group or aryl group, to produce an optically active 4-halo-3-hydroxybutyric acid ester or a preparation thereof is permitted to act on said 4-halo-acetoacetic acid ester and the product optically active 4-halo-3-hydroxybutyric acid ester is harvested. Thus, the desired optically active 4-halo-3-hydroxybutyric acid ester of high optical purity can

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EUROPEAN SEARCH REPORT

Application Number
EP 94 10 0369

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Place of search THE HAGUE		Date of completion of the search 28 April 1995	Examiner Delanghe, L
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons * : number of the same patent family, corresponding document			

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Application Number
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